REPORT OF THE AD HOC GROUP ON THE INTERNATIONAL REDFISH SURVEY IN THE NORWEGIAN SEA

| 1

16–18 SEPTEMBER 2008, COPENHAGEN, DENMARK, REVISED JANUARY 2009

1 Executive summary

In August 2008, Norway, the Russian Federation and the Faroe Islands conducted a joint survey on beaked redfish (*Sebastes mentella*) in the Norwegian Sea. The objectives of the survey, as set by NEAFC-AM 2007/58 were to measure the horizontal and vertical stock distribution and provide an abundance estimate. During the two weeks of investigation, the distribution, abundance and biology of *S. mentella* in the Norwegian Sea were studied by means of hydroacoustics and pelagic trawling onboard three commercial vessels: Atlantic Star (Norway), Osveyskoe (Russia) and Skálaberg (Faroes).

S. mentella was observed between 100 and 800m, with maximum concentrations in the 350-550m depth layer. This depth corresponds to the Deep Scattering Layer (DSL), were high concentration of small preys occur (myctophids, shrimps, cephalopods,...). *S. mentella* was observed in more than 90% of the trawls, over most of the area covered by the survey. The geographical distribution of the stock extended beyond the survey coverage, so only a fraction of the population could be studied by the survey. Generally larger and older individuals were found in the southern area of the study. More than 90% of individuals were older than 15 years, supporting the observed very low recruitment in the recent decades.

At the time of the meeting, not all hydroacoustic data was made available to the group. In addition there were important methodological difficulties and discrepancies which will require additional work to be resolved. For these reasons, a joint robust estimate of distribution and abundance of *S. mentella* in the Norwegian Sea can not be produced at the time of writing this report.

A series of recommendations are proposed to further analyse the data collected during the survey and converge towards common methodologies in future surveys. It is planned that an updated analysis of the data collected during the survey will be produced in early 2009.

2 Introduction

The meeting opened on the 16th September 2008 at 9:00 at ICES headquarters in Copenhagen. Christoph Stransky (Germany) was elected chairman for the meeting. There was a brief introduction by the chairman on security issues in the ICES building and the use of the ICES internet sharepoint system. Although the meeting is not a formal ICES meeting, it was agreed that the documents and report of the meeting would be placed on the ICES/SGRS sharepoint.

2.1 Objective of the meeting

Following the request from NEAFC (AM 2007/58), Norway, Russia and the Faroe Islands conducted a joint survey on redfish in the Norwegian Sea during August 2008. The objective of the meeting was to report on the international survey on *Sebastes mentella* in the Norwegian Sea in August 2008 and to provide relevant information on horizontal and vertical distribution and abundance.

2.2 Adoption of the Agenda

The agenda (annex 2) was adopted.

2.3 Participants

There were seven participants from 5 countries:

Eckhard Bethke (Germany), Kristján Kristinsson (Iceland), Andrey Pedchenko (Russia), Benjamin Planque (Norway), Jákup Reinert (Faroes), Fróði Skúvadal (Faroes), Christoph Stransky (Germany).

The detailed contacts for participants are given in Annex 1.

The grouped missed the expertise of some of the survey participants who had been collecting and scrutinizing the hydroacoustic data. This resulted in difficulties in resolving important issues linked to the analysis of hydroacoustic data. This is further discussed in the report and a recommendation is provided to address the issue (Annex 3, Rec. 7).

2.4 Structure of the report

The report is divided into three main sections. The first section presents the sampling methodology used by the three vessels for hydroacoustics and biological sampling. The second section reports the results available at the time of the group meeting. The third section discusses the results with regards to methodological aspects and provides a series of recommendations for improved planning, coordination and harmonization of the survey protocols in the future.

3 International survey on beaked redfish (*S. mentella*) in the Norwegian Sea in August 2008

3.1 Material and method

3.1.1 Vessels timing and survey area (Table 1, Figure 1)

<u>Norway</u>: the survey was conducted onboard the commercial trawler Atlantic Star. The survey extent and sampling plan was done according to the survey planning document (Annex 5) except for the following:

- Transect 10 was sampled before transect 9 (this was done so that the Atlantic Star could still meet with F/T Osveyskoe, because of the late departure of the latter, see below),
- The total number of trawl hauls was reduced to 24, to fit with available time during the survey. This resulted in 72 biological samples (3 samples per trawl haul when using the multisampler cod-end),

- Transects 13 and 14 were shifted slightly westward to cover a larger fraction of the international waters and less of the shelf break area.

<u>Russia</u>: the Russian part of survey was carried out by PINRO, the Knipovich Polar Research Institute of Marine Fisheries and Oceanography in Murmansk, with the commercial trawler "Osveyskoe" according to the survey planning document (Annex 5). However, some of the planned researches were altered and rescheduled prior to and during the survey:

- The survey started later than planned, on the 15th August after calibration of acoustic equipment near Tromsø. The survey ended on the 28th August (14 days in total).
- On some parts of the transects, trawling was not possible and only hydroacoustics registrations were performed. This was due to the vessel awaiting the permission by Russian authorities to fish in Norwegian waters (tracks 7 and 8) or adverse weather conditions (tracks 5 and 7).

<u>Faroe Islands</u>: the survey was conducted onboard the commercial trawler Skálaberg. The survey extent and sampling plan was done according to the survey planning (Annex 5) except for the following:

- Late start (14th August) due to preparation of vessel and fishing gear. This was
 the reason for why only 23 trawl stations were conducted in contrast to the 32
 originally planned.
- Tracks were followed according to the original planning except for a slight shortening of transects 1, 3, 4 and 5 in order to meet up with the Russian vessel Osveyskoe.
- Intercomparison of the hydroacoustic registrations between Skálaberg and Osveyskoe were not performed due to bad weather conditions. Instead two parallel trawl hauls were done, on 300 and 400 m headrope depth, respectively.

The extent of the survey and the sampling locations (trawls and acoustics) of the tree vessels is provided in Figure 1 and Table 1.

3.1.2 Biological sampling (Tables 2 and 3)

Norway: biological sampling onboard the Atlantic was carried out using the Gloria Trawl 2048/HO (100m x 100m opening) fitted with a multiple cod-end sampling device: the multisampler. This allowed for successive sampling at three distinct depth within one trawl haul and without contamination from one depth to the next and no sampling during shooting or heaving of the trawl. The sampling was carried out following the agreed recommendations set in the survey planning document. However, some adjustments were made for practical reasons. These are listed below:

- When the total number of fish of a given species was too large, the total number of fish was estimated from the total catch weight and the mean individual weight from a sub-sample (typically 100 fish).
- If the catch was so large that not all fish from one species could be weighted on the small scale, the total quantity was assessed by multiplying the number of fish baskets by the average weight of a basket (from a subsample).

- When using the multisampler, each trawl haul results in three biological catches.
 Each catch was treated separately. Otoliths were sampled from the first 10 individuals.
 Weight, sex, and maturity were determined for the first 33 individuals.
 Length and parasite infestation were recorded for the first 100 individuals. There was no recording of pigmentation or stomach content.
- Genetics sampling was only performed on 5 catches. Up to 100 individuals were sampled (fin clips, otoliths, length, weight, sex and maturity).

<u>Russia</u>: biological sampling onboard "Osveyskoe" was carried out using the Gloria Trawl 2048 (Table 2). The sampling was carried out following the agreed recommendations set in the survey planning document (Annex 5). The following adjustments were made:

- Total catch was weighted in each trawl, even when large quantities of fish were caught.
- Stomach fullness as well as parasite infestation, pigment patches and muscular melanosis were recorded according to the method described in Bakay and Karasev (2001). Analysis on the stomach contents were done for 576 individuals (mostly within DSL) and parasite for 1575 individuals.
- The fin clips and otoliths were sampled according PINRO method of biological sampling.
- Genetics sampling was performed on 11 catches (585 individuals).

<u>Faroe Islands</u>: the biological sampling onboard the Skálaberg was conducted using the Red Lion trawl 3072/HO (130m x 170 m) for the first 18 stations and the Gloria 4096/HO (200 m x 200m) for the remaining 5 stations. Both cod-ends were fitted with a 40 mm mesh 12 m long piece of netting in the hind most part. The biological sampling was followed performed according to the agreed recommendations. However there were some deviations from the plan for practical reasons and these are listed below:

- When the catch of redfish was too large to be weighted on the small-scale weight, the total catch was calculated from processed weight + sampled weight. The trawlers make use of a conversion factor of 1.7 to calculate the round weight from the processed fish. This factor was used to calculate the weight of round fish processed.
- On average, 100 *Sebastes mentella* were randomly sampled for weight, length, sex and maturity except from station 5 where 205 fish were sampled.
- Due to lack of personnel the genetic sampling was limited to 30 fish per station on fifteen stations scattered over the survey area.
- Parasite and pigment recordings was done on the fish that were sampled for otoliths, i.e. up to 30 *Sebastes mentella* per station were sampled.
- All species other than *S. mentella* were recorded as random subsamples taken from the total catch. The proportion of other species than *S. mentella* in the total catch was calculated as the proportion in the in the subsample relative to the total catch.

All length measurements were done at the cm below.

The trawl characteristics and biological sampling for the three vessels are summarized in tables 2 and 3.

3.1.3 Hydroacoustic sampling (Table 4)

Norway: hydroacoustics was conducted following the recommendations from the survey planning document (Annex 5). The hydroacoustics calibration was conducted on the first day of the survey. Results were highly consistent with previous calibrations conducted in May 2008 and August 2007 suggesting very minor drift in the instrument. Vessel noise measurements were conducted on the last day of the survey in calm weather conditions. The max recorded noise did not exceed -132dB for vessel speed between 7 and 13 knots. Overall the weather conditions during the survey were calm, allowing for 'clean' hydroacoustic registrations throughout the area surveyed. The registrations were recorded down to 1000m depth (instead of the 750m originally planned).

Scrutinizing of the acoustics data was done by echo-integration using the LSSS software. Contrary to the Irminger Sea, redfish in the Norwegian Sea is found mostly within the DSL, and in smaller quantities above (mixed with other fish species) and below (almost pure). The echo-integration was therefore performed with Sv thresholding to remove low-energy echoes which results from smaller targets in the DSL. Integration was done in a series of depth layers selected on the basis of vertical structures visible on the echogram and the information for the nearest trawl catches. In each layer, the threshold was raised up to a level where the DSL (or other 'background' layer) could not be seen. This was often around -72dB. The sA was then allocated to fish targets and divided between fish species according to sa proportions in the nearest trawl hauls (sa proportions are directly provided by the 'trawl module' of LSSS on the basis of species quantities and length distribution in the catch). The threshold was then brought back to -82dB and the additional sA was allocated to the category 'plankton'. In the upper 100m there was no trawl sampling so all energy was allocated to 'plankton' and 'other'. The data was not scrutinized below 800m. The scrutinizing was performed by 5nm blocks and the data exported into report files with a resolution of 0.1nm and 10m depth. Prior to abundance estimation, possible outliers were removed by filtering out the sa values that were more than 5 times greater than the median sA in the surrounding 0.5 nm. As fish below 600m had a significantly greater length than those above, two different target strength (TS) values were used in the abundance estimate: one for the layer 100-600m and one for the layer 600-800m.

Russia: before carrying out of shooting calibration on reference sphere about island Vannøya (Troms) on depth of 25 m has been executed.

Hydroacoustic sampling was carried out using echosounder ER60 (version 2.1.2) on frequency 38 kHz and systems of postprocessor processing FAMAS and BI60. Echointegration during trawling about 750 m were conducted. The method consists in calculation of amount of individual fishes on echograms (for an interval of 1 nautical mile), average force of the purpose $\langle TS \rangle$ which exceeds certain value TS_{min} with the further estimation S_{Aac} for these fishes or under classical formulas of hydroacoustics, or on the algorithms received experimentally. For example, special measurements have shown, that for the redfish in length 36 - 40 cm ($\langle TS \rangle$ = - (40-39) dB) on depth of 600-700 m the amount of fishes on mile of a way n=4 is approximately equivalent S_{Aac} = 1 m² / nm².

Identification by acoustic way of the redfish settling down below edge of DSL, had experimental character and was possible only in case of satisfactory "noise" conditions of environment (for example, at sweeps or at favourable weather).

For a trawl Gloria the coefficient of trawl efficiency has been accepted equal K=0.250 and the formula of calculation s_A for the redfish for the period sweep in DSL and under it looked like s_A = K*Catch (Results from observations in the Irminger Sea in 2007).

Calculation of number and a biomass of *S. mentella* have been averaged over 5 nm intervals. Acoustic backscattering coefficients (sA) of *S. mentella* in each 5 nm block were estimated separately above the DSL (as a rule, is higher 300-400 M), and within/below the DSL.

Faroe Islands: hydroacoustics was conducted following the agreed recommendations from the planning document. The Skálaberg had recently installed the EK 60 and split beam transducer. The hyrdoacoustic calibration was conducted on the 13th of August. The results from the calibration showed satisfying results and were done according to the EK 60 manual. Vessel noise measurements were done on the 15th of August. Some scrutinizing of the acoustic data were done onboard. The energy was allocated to species according to the nearest trawl haul. The acoustic data are not fully processed yet but are expected to be ready for the acoustic workshop in Tromsø on 24-26 Nov. 2008.

3.1.4 Hydrographic measurements (Table 5)

Norway: hydrographic measurements were done using a CTD probe SAIV AS/SD204 calibrated prior to the survey. The probe was attached to the multisampler and recorded temperature and salinity every 10 seconds. The maximum depth of hydrographic measures was set by the maximum trawling depth. During trawling, the multisampler-CTD was usually 40 to 70m below the headline depth.

Russia: hydrographic measurements were done using the Simrad temperature sensor of FS20 attached on trawl headrope. During the survey in a few different points the data of sensor was calibrated against data from a CTD probe FSI NXIC. After the survey, temperature data from the FS20 sensor was corrected and used for analyse. The maximum depth of temperature measures was set by the maximum trawling depth (headrope) and was about 600 m.

<u>Faroe Islands</u>: hydrographic measurements were done using Star-Oddi CTD Data Storage Tags attached to the trawl. Two tags were attached to the trawl at each station, one on the headline and one on the groundline. The tags were set to measure temperature, depth and salinity every second. The results were then after each haul read from the tags and an average temperature derived from the data.

3.2 Results

3.2.1 Species composition (table 6)

Sebastes mentella was the most common species found and occurred on average in 93% of the trawl samples (note that most trawls were performed around the depth of maximum density of *S. mentella*).

Blue whiting (*Micromesistius poutassou*) was also commonly found (83% occurence) above and within the DSL. The non-commercial species 'ribbon barracudina' (*Arctozenus risso*) also occurred throughout the sampling region (74% occurence).

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Catches of other species included small individuals, mostly in the DSL, such as shrimps and myctophids (30%). Cornish blackfish was also commonly found in the middle and southern regions (23% occurence).

Other fish species included saithe (29%), herring (24%), greater argentine (13%), golden redfish (6%), mackerel (8%), cod (7%), Greenland halibut (5%), and haddock (1%).

3.2.2 Horizontal and vertical distribution

Norway: because trawls were performed at different depths, sometimes at the depth of maximum density of redfish and sometimes in much shallower or deeper waters, it is not possible to derive a reliable horizontal distribution from the trawl data alone. Therefore, the horizontal distribution is described here on the basis of hydroacoustics registrations. *S. mentella* was registered in almost all the area covered (Figure 96), except for the north-westernmost part where water temperature was the lowest (Figure 118). The highest densities were found in international waters and close to the shelf break around the 600m isobaths.

The vertical distribution of S. mentella was studied from the trawl catch and the hydracoustic registrations. The vertical distribution of catch rates (Figure 7) indicate that S. mentella was found at all depths. 90% of the catches were distributed between 200 and 600m and 50% of the catches were taken from the layer 400-550m, mostly within the DSL (see Figure 85 for examples of DSL in acoustic registrations). Although hydroacoustics is limited at depth because of energy spreading (and consequent loss of signal from individual fish targets), the vertical distribution of area backscattering coefficients (sA) is very close to that of trawl catch rates. The shift between cumulated distributions of catch rate and sA against depth indicate that hydroacoustic registrations are biased toward higher abundances in shallower waters (which is to be expected). However, the difference between the distribution obtains from trawling and hydroacoustics is small which indicate that hydroacoustics registrations are probably valid throughout the water column, down to 800m. There are two main sources of bias associated with echo-integration in the DSL and at depth. The first is related to the misallocation of DSL targets to redfish (which results in an over-estimate of redfish abundance in the DSL). This can be partly resolved by sv thresholding in order to remove low energy targets (i.e. plankton and small fish) and retain high energy ones (e.g. redfish). This may lead to the second type of bias which is due to the reduction in the effective beam angle as a result of sv thresholding (which results in an underestimate of redfish abundance at great depths). These are further discussed in the discussion section (3.3)

<u>Russia</u>: The analysis of echograms has shown, that in the daytime rarefied concentrations of *S. mentella* as of 200-350 m were usually distributed on the depths, mixing up with DSL, blue whiting, ribbon barracudina (*Arctozenus risso*). With approach of darkness the top border of DSL rose up to depths of 50-100 m and identification of concentrations of *S. mentella* by acoustic way became inconvenient and to allocate values sa for the redfish on background of DSL somewhat it was possible to allocate with change of a threshold at postprocessor processing on BI60 or FAMAS. Control sweep in the daytime showed catches from 9 to 550 kg per hour trawling.

<u>Faroe Islands</u>: in the western-most part of the survey area there were no *S. mentella* catches. This was particular evident when getting near the Icelandic continental shelf. In this area there was also a cold front with temperatures as low as 1°C at 400 m depth (Figure <u>118</u>). Apart from these stations *S. mentella* was caught on all stations

with the highest catches in the Northern part of the Faroese EEZ and in the Southern part of the International waters. The highest catches were in the DSL at 350 m at temperatures of 3-4°C.

3.2.3 Assessment by trawl and acoustics method

Under certain assumptions, and following specific methodologies, it is possible to estimate the total abundance of redfish in the surveyed area using trawl catches and/or hydroacoustic registrations. The group tried the two methods but this was done with limited success due to uncertainties in some assumptions and lack of common agreed methodologies. The results from these calculations are presented below.

3.2.3.1 Trawl abundance estimates

The trawl method is the simplest. The 2 most important assumptions are 1) that the trawl samples are representative of the population in the area (i.e. sufficient trawl hauls in different sub areas and depth strata and sufficient sampling volume) and 2) that catchability of redfish in the trawl is known. Assumption 2 is unknown and the group could only perform calculations with an assumed catchability of 100%. Furthermore, 3 different types of trawls were used during the survey and the degree to which data from different trawls can be compared is unknown.

The trawl based calculations are therefore done in an indicative manner but can not provide a robust abundance estimate at the present time.

The procedure for trawl based abundance estimates is as follows: all trawl samples are grouped by depth strata (50m for Norwegian data and 100m for Russian and Faroese data) ranging from 100 to 800m. For each stratum the mean catch rate (by nm² and 100m depth) is calculated. The sum of the catch rate (i.e. catchrate per nm² for the whole water column) is then multiplied by the survey area to provide a total abundance.

For the north part of the survey (Norway) the total area sampled is 53,720 nm². The mean catch rate is 3.8 tonnes/nm². The estimated total abundance is 203,000 tonnes.

For the middle part of the survey (Russia) the total area sampled is 97,865 nm². The mean catch rate is 2.8 tonnes/nm². The estimated total abundance is 276,000 tonnes.

For the southern part of the survey (Faroes), the total area sampled is $73,140 \text{ nm}^2$. The mean catch rate is 0.9 tonnes/nm^2 . The estimated total abundance is 65,000 tonnes.

3.2.3.2 Hydroacoustics abundance estimates

The hydroacoustics estimates were performed differently for the three parts of the survey and the individual procedures are detailed below.

Norway: the procedure for hydroacoustics abundance estimate is described in section 3.1.3. On average, the depth integrated backscattering coefficient (s_A) value was $33m^2/nm^2$. The TS value used for *S. mentella* was based on the length dependent equation TS=20log*L-k*, with *k*=68. This value is based on recent estimates (Gauthier and Rose, 2001, 2002) and measurements based on individual fishes (Kang and Hwang, 2003). However, there is still an ongoing debate on whether *k* should be set to 68 or 71.3 (as currently used in the Irminger Sea). The effect of setting *k* to 71.3 instead of 68 is to raise the hydroacoustics abundance estimate by a factor of 2. Further investigations on the *in situ* target strength of *S. mentella* need to be conducted to resolve this issue.

The total abundance estimate, over the 53720nm² area is estimated to 395,000 tonnes. This abundance estimate is significantly larger than the value obtained from the trawl estimate (203,000 tonnes). This may result primarily from under estimation in the trawl estimates due to the true catchability being less than 100%. It may also result from misallocation of acoustic energy from small targets in the DSL to *S. mentella*. These issues would need to be further investigated.

<u>Russia</u>: the estimation of redfish density distribution was carried out via the trawling method by means of recalculation of sizes the redfish catches in acoustic units saturation application of the program "Severer".

Calculation of number and biomass of the redfish was carried out by the stratified method, i.e. breakdown of all research area on spatial strata, the size 2° longitudes on 1° breadths. The equation for the redfish TS=20LgL-71.3, recommended SGRS for estimation stock abundance of the redfish in the Irminger Sea in 2007, was used. According our calculation the biomass of the redfish in our part of Norwegian Sea was at 2.14 time less when using equation TS=20LgL-68,0. Total number and a biomass of fishes on the area was determined as sum total of fishes from separate rectangular.

The total abundance estimate of the redfish on the area $97,865 \text{ nm}^2$ is estimated about 76,700 tonnes ($127 \times 10^9 \text{ individuals}$). The abundance estimate above DSL is 24,600 tonnes ($41 \times 10^9 \text{ individuals}$), and in the DSL and deeper is 52,100 tonnes ($86 \times 10^9 \text{ individuals}$).

<u>Faroe Islands</u>: the analysis of hydroacoutics data from the Faroese part was not completed at the time of the group meeting (section 3.1.3). The final scrutinizing and allocation of the acoustic energy to different organisms will be done before the hydroacoustic scrutinizing workshop planned in Tromsø in late November 2008.

3.2.4 Biological data

Length, sex, maturity, parasites and pigmentation data were compiled for the three areas. For length and sex, the data were also analysed by depth strata in the northern part of the survey (Norway).

The lengths of individuals sampled ranged from 29 to 46 cm (Figure 3). The mean length increased towards southern areas with 36.6 cm in the north (Norway), 37.0 cm in the middle (Russia) and 37.7 cm in the south (Faroes). The mean length of females was greater than that of males by one cm or more in all areas.

The length distribution also varied with depth of sampling (Figure 4, northern area). Individuals collected in the layer 100-300m were slightly longer (mean 36.8 cm) than those found in the layer 300-600m (mean 36.3 cm). Below 600m, the mean recorded length was much greater: 38.7 cm.

The age distribution was determined for the Norwegian and Faroes data, from otolith reading carried out at the Institute of Marine Research (Norway) following the protocol recommended by the ICES Workshop on Age determination of Redfish (Nanaimo, Canada, September 2008). In the Northern area, 90% of the population is composed of fish older than 15 years and there were no individuals younger than 10 years (Figure 5). The age distribution of males and females is almost identical. On the other hand, there are important differences in age structure in different depth strata. The layer 300-600m where most fish are found is dominated by individuals around 18 years. In the above layer (100-300m) maximum densities are found for fish of 19 and 27 years. In the deeper layer (600-800m) maximum densities are found for fish of

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17 and 27-31 years. Overall the proportion of older individuals is greater in the deep layer. The length-at-age for females are significantly greater than for males (Figure 6)

In the southern area, the proportion of old individuals is much higher with 50% of the individuals older than 33 years and 90% older than 19 years. The maximum density is at 39 years (Figure 7). Age reading from the Norwegian and Faroese samples reveal that: 1) population age structure is depth dependent and region dependent, 2) that most fish are older than 15 years, confirming the existing evidence for low recruitment in the past 15 years, 3) that females are larger than males, due to differential growth rates.

The sex ratio was in favour of females in the northern part of the survey area, but males dominated in the middle and southern areas (Table 7). An analysis of sex ratio by depth strata in the northern part shows that the proportion of females is greater in the layer 100-300m (61%) and even greater in waters deeper than 600m (72%) than in the DSL (52%).

Norway and Faroes recorded most individuals at a maturity stage of 2 (ICES scale for Norway and closely related Faroese scale for the Faroes). Russian maturity reading reported a majority of male stage 2 and female stage 3 (according to the ICES scale). Some discrepancies remain in the maturity scales used and in the reading of maturity by the different participants. These will need to be further investigated.

The stomach contents were analysed only for the central part of the survey area (Russia). In both the shallower and deeper layer, the majority of the redfish stomachs (98.6% shallow, 93.7% deep layer) were everted. The data of redfish catch within the DSL show that in stomach content was mostly constituted by small plankton (62%), shrimps (21%) and fish items (15%).

3.2.5 Hydrography

The oceanographic regime in the investigation area depends on inflow of warm and saline Atlantic water into Norwegian Sea from the North Atlantic Ocean (Figure 129). The Atlantic current transports warm water northwards, with the result that there are important for fish resources distribution in the Norwegian Sea.

While warm Atlantic water flows north along the coast of Norway, cold Arctic water flows southwards along the eastern coast of Greenland. Cold water from the Polar Basin flows through the Fram Strait. A part of it continues south through the Denmark Strait, while some passes into the Greenland Sea, the Icelandic Sea and the Norwegian Sea, producing a front between a warm eastern side and a cold western side. This front plays a central role in the distribution of species in the sea.

The peculiarities of temperature condition on the research area in August 2008 are illustrated by the horizontal distributions of temperature on 200 and 400 m depth (Figure 8). The main redfish concentrations were recorded along the gradient zone between Atlantic and Arctic waters southeasterly of the Mohn Ridge and in the central part of area. The temperature in the place of the greatest catch of redfish was: in the northern part within depth 350-600 m about 2.5-4.5 °C; in the central part within depth 400-600 m about 2.1-5.0 °C; in the southern part area within depth 300-450 m from 0.5 to 4.5 °C.

3.3 Discussion

3.3.1 General aspects

It is the first year that an international survey on *S. mentella* in the Norwegian Sea is conducted. The planning and coordination have been done within a limited time frame and without a proper planning structure (such as an ICES planning group). Five parties originally agreed for this survey to be conducted, but in 2008, it was only possible for three of them to conduct the survey (Norway, Russia and the Faroe Islands). Due to funding constraints, the survey was run from commercial trawlers rather than research vessels. All these factors resulted in non-optimal conditions for conducting the planned scientific research on redfish distribution and abundance. In addition, observation and data processing methodologies were not always harmonised to a sufficient level, making it difficult to combine the results from the three parties. All these limitations will need to be carefully considered and the necessary adjustment be made for future surveys.

3.3.2 Distribution and acoustic abundance estimate

The survey did not reach the boundaries of the spatial distribution of *S. mentella*. Future surveys will need to consider spatial expansion at least in the northern and western area. This will require at least one additional vessel, given a similar density of acoustic tracks and trawls as in the 2008 survey.

In the eastern and north-eastern part of the survey, the open-ocean component of the population overlaps with the demersal component. In particular over the shelf break area between the Barents Sea and the Norwegian Sea. This 'mixed' component needs to be considered. This will require dedicated observation and sampling methodologies, as the method used for the open-ocean (acoustics and pelagic trawling) is not adequate in this area. In addition, the stock identity of *S. mentella* in the North-east Arctic needs to be investigated further.

The *in situ* target strength of *S. mentella* is still a matter of debate. Recent bibliography and rapid calculations based on individual target counting and s_A estimates during the survey suggest that length dependent TS coefficient is about -68. However, in the Irminger Sea, this coefficient is usually set to -71.3. Such difference in the coefficient can alter abundance estimates by a factor of 2. It is therefore critical that dedicated research is conducted to determine unambiguously the in-situ target strength of red-fish in the Norwegian Sea.

The vertical distribution of *S. mentella* in the Norwegian Sea is different from the one generally observed in the Irminger Sea. A dominant feature is the presence of highest densities of *S. mentella* within the Deep Scattering Layer (DSL). Above the DSL, *S. mentella* is often found mixed with other fish species (blue whiting, herring,...) whilst it is found 'pure' below the DSL. The vertical distribution extends down to 800m. For these reasons, the methods to assess the distribution and abundance of *S. mentella* in the Norwegian Sea can not be transposed directly from those developed and used in the Irminger Sea. In particular, the hydroacoustics scrutinizing procedures and the trawl-acoustic regression models will need to be evaluated carefully.

The important discrepancies between s_A values from the Russian and Norwegian vessels can not result only from differences in abundance of *S. mentella*. The vessels acoustic performance and the methods used by the two parties to scrutinize the hydroacoustic registrations are different. How this affects the abundance estimate needs to be addressed and resolved. As there were no Faroese data presented at the time of

the meeting, it was not possible to compare these data with the two other vessels, but a similar effort of harmonization should be thought. In summary, tThere is an urgent need for harmonised methodology and onboard scrutinizing of the data, if a consensus view on horizontal distribution and abundance is to be reached. A first attempt to explore the discrepancies in scrutinizing methodologies was carried out during the hydroacoustic scrutinizing workshop held in Tromsø, Norway in November 2008. Much work remains to be done for robust hydroacoustic estimates to be obtained for the Norwegian Sea area. A first step in this direction in proposed in the form of an hydroacoustic scrutinizing workshop (recommendation 7, Annex 3).

3.3.3 Abundance estimate from trawl

The survey plan specifically stated that the abundance and spatial distribution would be studied by hydroacoustics, but because of the current difficulties and uncertainties with hydroacoustics estimates, a trawl based abundance estimate was attempted. However, there are large differences in gear (three different trawls) and depth zones between vessels, so only data by individual vessels can be presented. A robust abundance estimate from trawl would require standardisation of trawl equipment, standardisation of vertical distribution of trawl hauls and known catchability. This will be difficult to achieve with commercial vessels and would require dedicated research on pelagic trawl catchability of *S. mentella*.

3.3.4 Biological sampling and data processing

The vertical distribution of *S. mentella* is difficult to study with standard pelagic trawls which have a wide opening and require substantial time for shooting and hauling. The use of the mutlisampler by Norway proved to be an efficient way to assess the depth distribution of *S. mentella* without increase in the sampling time. The generalisation of the use of the mutlisampler should be thought of.

The sampling protocols for genetics and otoliths need to be better harmonised:

- The sampling for genetics was carried out differently onboard the three vessels. Whilst Norway collected up to 100 fish in only 5 stations, Russia collected 585 individuals in 11 stations and the Faroes collected up to 30 fish in 15 stations. Harmonisation of genetics sampling protocols in critical for the subsequent analysis of samples with micro-satellites or other methods.
- At present, Norway and the Faroe Islands have followed a random sampling protocol for otoliths, whilst Russia is sampling following a length stratified sampling scheme. The two approaches yield slightly different results in age-structure estimates. They will need to be assessed and a common sampling procedure selected.

In Norway, otolith reading is currently performed using the most recently internationally agreed protocols (ICES, 2008). The Faroe Islands do not hold the technical capacity to analyse the otoliths collected in 2008. <u>However, Faroese otoliths were sent to Norway and age reading performed in the same way.</u> Cross-comparison of reading between Norway and Russia should be carried out. Training of Faroese and Russian readers to internationally agreed methods is also needed. At the time of the meeting no age reading was available. Some results will be available in January 2009.

The analysis of parasites and pigmentation was fully carried out by Russia but only partially by Norway and the Faroe Islands. Harmonisation of the sampling protocols and observation methods should be done through a common training workshop.

3.3.5 Hydrography

During 2008, hydrography measurements were done on an ad hoc basis, considering the constraints of individual commercial vessels (winches), available hydrographic equipment and time. The three vessels used instrument attached to the trawl (headline or multisampler) but with different precision, accuracy and data acquisition frequencies. Temperature and salinity measurements made to acceptable hydrographical standards can only be achieved with high precision instruments, calibration, water sampling and vertical stations. This will require additional time and equipment if conducted in the future. Alternatively, measurements taken from trawl attached instruments may provide acceptable data for redfish habitat identification and water masses characterisation at the scale of the survey. The precision, accuracy and measurement protocols for temperature and salinity will need to be discussed ad harmonised for future surveys.

4 Future surveys, 2009 and beyond

The group agreed that a second international survey on *S. mentella* in the Norwegian Sea should be conducted in 2009. Many of the sampling and data processing methodological issues should be clarified during the planning of this second survey. This can be achieved through a series of workshops (see recommendations 3, 7, 8, 9 in Annex 3) and by the constitution of a dedicated planning group under the auspices of ICES.

The current survey effort is not sufficient to cover the geographical distribution of *S. mentella* and an increase in the number of vessels and/or survey duration will be needed. Optimally, the survey should be conducted from research vessels.

From 2010 onwards, a regular international Norwegian Sea survey could be conducted every two years, in alternation with the international Irminger Sea survey currently planned and reported by ICES/SGRS (Study Group on Redfish Stocks).

5 Acknowledgements

The group expresses their thanks to the International Council for the Exploration of the Sea for hosting the meeting in its headquarters in Copenhagen and providing all necessary help for the conduction of the meeting.

6 References

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7 Tables

Table 1. Extent and coverage of the survey

Country	Norway	Russia	Faroe Islands	
Vessel	Atlantic Star	Osveyskoe	Skálaberg	
Days in the field	14	13	12	
Number of hauls	24 (x3)	28	23	
Min/max trawling depth	100 m / 800 m	100/600 m	200 m / 600 m	
Distance for acoustics registration	1,350 nm	2,110 nm	1,219 nm	
Area surveyed	53,720 nm ²	97,865 nm ²	73,140 nm ²	
Number of CTD casts	24	28	18	

Table 2. Trawl specifications

Country	Norway	Russia	Faroe Islands	
Manufacturer/ref	Hampidjan / Gloria 2048 HO	Hampidjan / Glo- ria 2048 HO	Vónin/Red Lion 3072 Hampidjan / Glo- ria 4096 H20	
Opening	100m	100m	130/195	
Width	100m	110m	168 /196	
Cod end	Multisampler (3 bags) / inner net 40mm	12-m inner net 40mm	12-m inner net 40mm	

Table 3. Summary of the biological sampling

Country	Norway	Russia	Faroe Islands
Total number / bio- mass of redfish caught	6,378 ind. / 3,892 kg	9,673 ind. / 6,105 kg	9,344 ind. / 6,401 kg.
Number of length measurements	2,914	6,116	1,848
Number of pairs of otoliths collected	940	1,225	584
Number of feeding analyses	-	576	-
Number of parasites analyses	-	1,175	-
Number of stations/individuals with genetics	5 / 410	11/ 585	15/ 431

Table 4. Instrument settings of the acoustic equipment onboard the participating vessels

Country	Norway	Russia	Faroe Islands
Vessel	Atlantic Star	Osveyskoe	Skálaberg
Echo/sounder/Integrator	Simrad EK60/LSSS	Simrad EK60/BI60/Famas	Simrad EK60/EchoView
Frequency	38kHz	38kHz	38kHz
Transmission Power	2000W	4000W	2000W
Absorption coefficient	9.65 dB/km	9.8 dB/km	9.8 dB/km
Pulse length	1.024	1.024	1.024
Bandwidth	2.43 kHz	2.43 kHz	2.43 kHz
Transducer type	ES 38-B	ES 38-B	ES 38 B
Two-way beam angle	-20.6 dB	-20.6 dB	-21.0 dB
Integration threshold	-82dB	-70 bB	-70dB
Sound speed	1493 m/s	1494 m/s	1494 m/s
Transducer gain Sv	25.59 dB	25.01 dB	
Transducer gain TS			25.41 dB

Table 5. Temperature and salinity sensor specifications

Country	Norway	Russia	Faroe Islands	
Manufacturer / ref	SAIV AS / SD204	Simrad/ tempera- ture sensor of FS20	Star-Oddi/DST CTD and DST milli	
Pressure resolution / accuracy	0.01dbar / 0.1dbar	0.1dbar / ±1%	0.03%/+/-0.4 %	
Temperature resolution/accuracy	0.001°C / 0.01°C	0,05 °C / 0,5 °C	0.032°C/0.1°C	
Salinity resolution/accuracy	0.01 /0 .015	-	0.02 (PSU)/ +/- 0.75 (PSU)	
Data acquisition frequency	0.1 Hz	-	1 Hz	
Type of profiling	CTD attached to the multisampler (cod-end)	Attached to headrope	CTD DST attached to the head line and fishing line	

Table 6. Species occurrence.

	Ti	rawls with spe	cies presen	t, Country (to	tal no. of tr	awls)	
Country	Norw	ay (72)	Russ	sia (28)	Faroe Is		
Species (Latin name)	Number	Percentage	Number	Percentage	Number	Percentage	(Avg. % for ranking)
Beaked redfish (Sebastes mentella)	69	96%	27	96%	20	87%	93%
Blue whiting (Mi- cromesistius pou- tassou)	48	67%	27	96%	20	87%	83%
Ribbon barracudina (Arctozenus risso)	45	63%	25	89%	16	70%	74%
Squid (Gonatus spp.)	29	40%	21	75%	-	-	38%
Myctophids (<i>Mycto-phidae</i>)	31	43%	6	21%	6	26%	30%
Saithe (<i>Pollachius</i> virens)	4	6%	12	43%	9	39%	29%
Herring (<i>Clupea</i> harengus)	20	28%	5	18%	6	26%	24%
Cornish blackfish (Schedophilus medusophagus)	2	3%	11	39%	6	26%	23%
Shrimp	1	1%	10	36%	2	9%	15%
Glass shrimp (Pa- laemonetes spp.)	28	39%	-	-	-	-	13%
Sagittal squid (<i>Ommastrephes</i> <i>sagittatus</i>)	-	-	-	-	3	13%	13%
Greater argentine (Argentina silus)	6	8%	7	25%	1	4%	13%
Mackerel (Scomber scombrus)	2	3%	1	4%	4	17%	8%
Cod (Gadus mor- hua)	4	6%	1	4%	3	13%	7%
Golden redfish (Se- bastes marinus)	-	-	4	14%	1	4%	6%
Greenland halibut (Reinhardtius hip-poglossoides)	10	14%	-	-	-	-	5%
Krill (Euphausiacea)	8	11%	-	-	-	-	4%
Atlantic pomfret (Brama brama)	-	-	-	-	2	9%	3%
Lumpsucker (Cyclopterus lum- pus)	6	8%	-	-	-	-	3%
Northern Wolffish (<i>Anarhichas den-</i> <i>ticulatus</i>)	-	-	-	-	1	4%	1%
Haddock (Melanogrammus aeglefinus)	-	-	1	4%	-	-	1%
Octopus (Octopus spp.)	1	1%	-	-	-	-	<1%
Northern Rockling (Ciliata septentrion-	1	1%	-	-	-	-	<1%

Table 7. Sex ratios for *S. mentella*

Country	Norway	Russia	Faroe Islands
%Males	45.5%	63.2%	61.6%
%Females	54.5%	36.8%	38.4%

Table 8: Infestation by parasites (copepod S. lumpi) and pigmentation for trawls above the DSL (top) and trawls within and below the DSL (below).

		Norway			Russia		F	aroe Island	ls
Trawls above DSL	males	females	total	males	females	total	males	females	total
External damages							•		
No. of fish examined	185	308	493	203	152	355	102	92	194
No. of fish with S.lumpi and/or rem- nants	37	63	100	128	97	225	9	5	14
% of fish with <i>S.lumpi</i> and/or remnants	20,0	20,4	20,3	63,1	63,8	63,4	8,8	5,4	7,2
No. of <i>S.lumpi</i> and/or remnants	54	108	162	301	278	579	10	6	16
Abundance index of S.lumpi invasion	0,29	0,35	0,33	1,5	1,8	1,6	0.09	0.07	0.08
No. of fish with ex- ternal pigment spots				6	4	10			
% of fish with exter- nal pigment spots				3,0	2,6	2,8			
Muscular melanosis							_		
No. of fish examined				203	152	355			
No. of fish with mus- cular melanosis				1	2	3			
% of fish with muscu- lar melanosis				0,5	1,3	0,9			
Trawls within and		Norway		Russia		Faroe Islands		s	
below DSL	males	females	total	males	females	total	males	females	total
External damages									
No. of fish examined	639	855	1494	514	306	820	198	192	390
No. of fish with S.lumpi and/or remnants	109	170	279	307	197	504	13	10	23
% of fish with <i>S.lumpi</i> and/or remnants	17,1	19,9	18,7	59,7	64,4	61,5	6,6	5,2	5,9
No. of <i>S.lumpi</i> and/or remnants	152	248	400	688	504	1192	18	12	30
Abundance index of S.lumpi invasion	0,24	0,29	0,27	1,3	1,6	1,5	0,09	0,06	0,08
No. of fish with ex- ternal pigment spots				9	16	25			
				9 1,8	16 5,2	25 3,0			
ternal pigment spots % of fish with exter-									
ternal pigment spots % of fish with external pigment spots									
ternal pigment spots % of fish with external pigment spots Muscular melanosis				1,8	5,2	3,0			

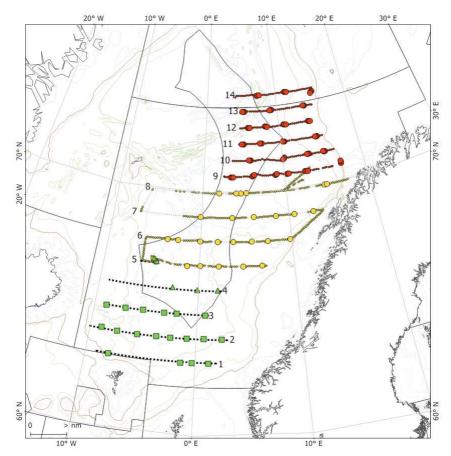


Figure 1. Geographical extent and sampling activity during the *S. mentella* survey in August 2008. Small dots show the location of 5nm sections retained for acoustic scrutinizing. Lager dots indicate the location of biological sampling (trawling) for Norway (red), Russia (yellow) and the Faroe Islands (green). Circles: Trawl Gloria 2048, triangle: Trawl Gloria 4096, squares: Trawl Red Lion. The acoustic data for the Faroese part is not available at the time of the report; acoustics tracks are thus shown as dotted lines.

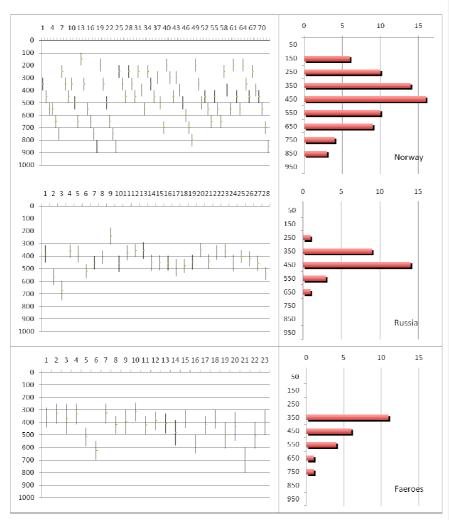
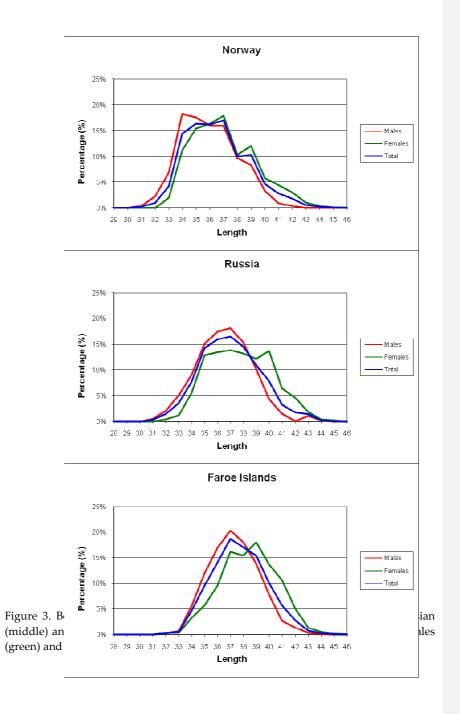


Figure 2. Vertical extent of trawling during the *S. mentella* survey in August 2008 for Norway (top), Russia (middle) and the Faroe Islands (bottom). The left panels display the sequence of trawls with vertical bars placed at the upper and lower limits of the trawl opening. The right panels display the frequency of trawls in 50m depth layers. The depth is calculated as the mean depth of trawling (not headrope depth, but depth between the depth of the headrope and the depth of headrope + opening).



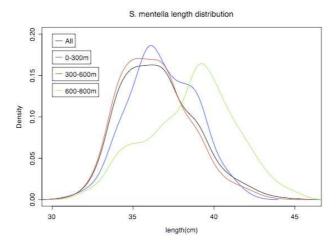


Figure 4. Body length distributions of S. mentella for three depth layers for the Northern part of the survey (Norwegian data only): layer 0-300m (blue), 300-600m (red) and 600-800m (green).

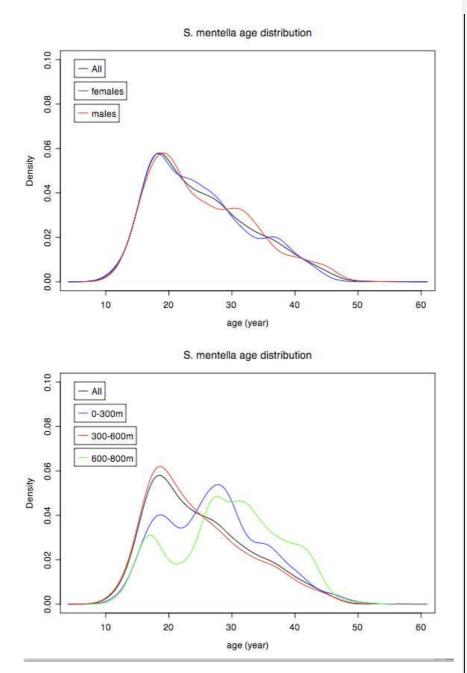
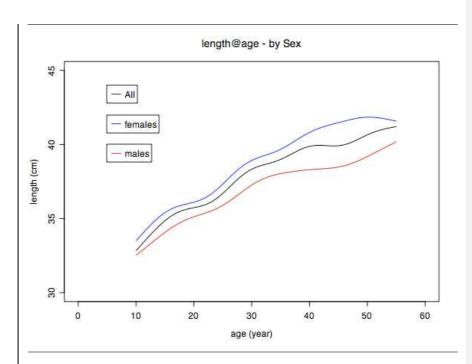
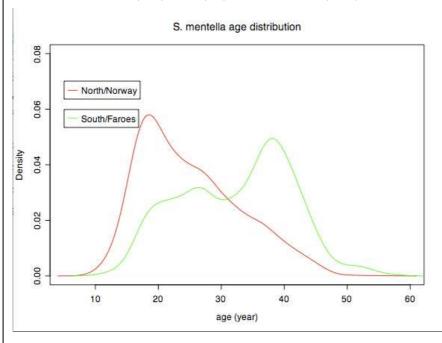


Figure 5. Age distribution of S. mentella in the northern part of the survey (Norway). Top: age distribution for females (blue), males (red) and sex-combined (black). Bottom: age distribution for the shallow (blue), middle (red), deep (green) or all depthcombined (black).



<u>Figure 6: length-at-age of *S. mentella* in the northern part of the survey (Norway) for females (blue), males (red) and sex-combined (black).</u>



<u>Figure 7: age distribution of *S. mentella* for the northern (Norway, red) and southern (Faroes, green) areas.</u>

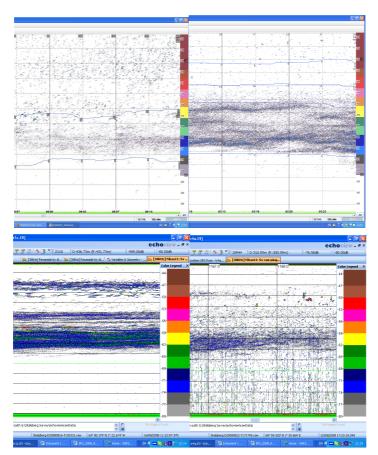


Figure §5. Examples of echograms recorded during the survey (top: Atlantic Star 100-800m, bottom: Skálaberg 0-700m). Common features include the Deep Scattering Layer (DSL) in diffuse (top-left) or dense (top-right and bottom-left) state and detection of individual targets (mostly redfish) above, within and below the DSL. Bottom-right echograms shows the effect of poor weather conditions on the background noise (most likely bubble attenuation).

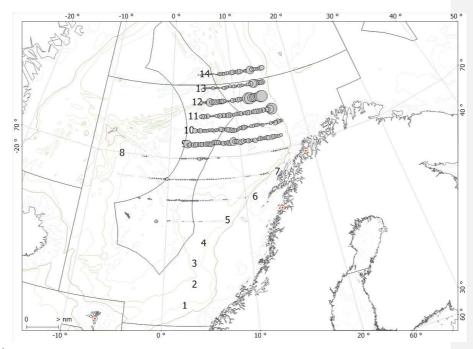


Figure <u>96</u>. Spatial distribution of area backscattering coefficient (sA) of *S. mentella* estimated for the Norwegian (north) and Russian (middle) part of the survey.

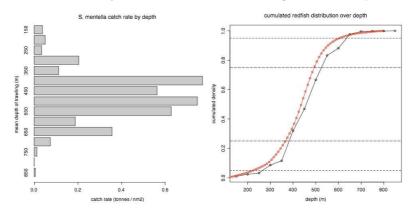


Figure <u>10</u>7. Left: Vertical distribution of catch rates. Right: cumulated density distribution of catch rates (black) and area backscattering coefficient (sa, red) as a function of depth. Dotted lines indicate the 5 and 95% probability levels. Dashed lines indicate the 25% and 75% probability levels. Data are from the northern part of the survey only (F/T Atlantic Star).

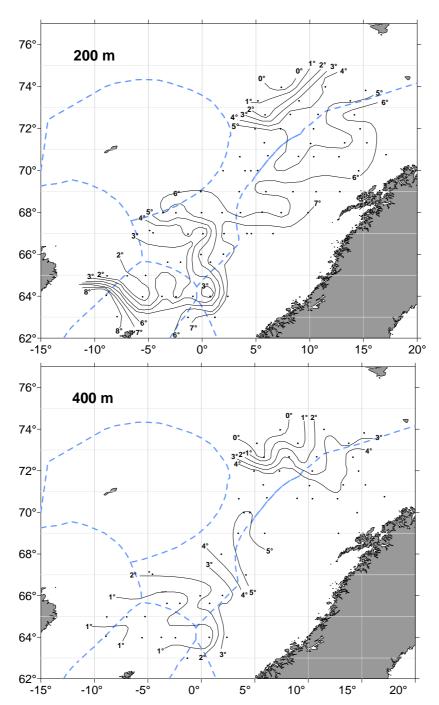


Figure 11-8. Horizontal distribution of temperature (°C) at 200m (top) and 400m (bottom). Black dots indicate the position of temperature measurements.

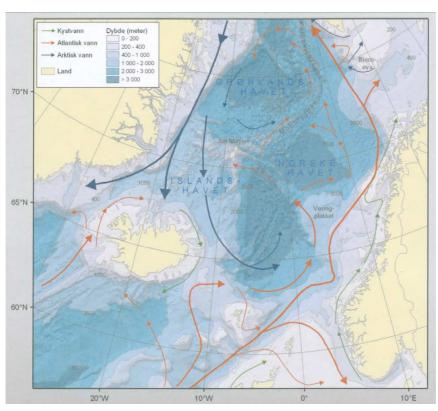


Figure <u>12</u>9. Horizontal distribution of the main currents in the Norwegian Sea. Red arrows: Atlantic waters. Blue arrows: Arctic waters. Green arrows: Coastal waters.

Annex 1: List of participants

Name	Address	Phone/Fax	Email
Eckhard Bethke	Johann Heinrich von Thünen Institute [vTI] - Federal Re- search Institute for Rural Areas, Forestry and Fisheries - Insti- tute of Sea Fisheries Palmaille 9, D-22767 Hamburg, Germany	Tel. +49 40 38905-203 Fax: +49 40 38905-263	eckhard.bethke@vti.bund.de
Kristján Kristinsson	Marine Research Institute, P.O. Box 1390 121 Reykjavik Iceland	+354 575 2000	krik@hafro.is
Andrey Pedchenko	Knipovich Polar Research Institute of Marine Fisheries and Oceanography (PINRO) 6 Knipovich st. Murmansk, 183763, Russia	+007 8152 473280 / +007 8152 473331	andy@pinro.ru
Benjamin Planque	Institute of Marine Research, Postboks 6404, 9294 Tromsø, Norway	+47 77 60 97 21	benjamin.planque@imr.no
Jákup Reinert	Faroese Fisheries Laboratory, Nóatún 1, P.O. Box 3051 FO-110 Tórshavn Faroe Is- lands	+298 353935/+298213092	jakupr@frs.fo
Fróði B. Skúvadal	Faroese Fisheries Laboratory, Nóatún 1, P.O. Box 3051 FO-110 Tór- shavn Faroe Islands	+298 353950	frodis@frs.fo
Christoph Stransky (chair)	Johann Heinrich von Thünen Institute [vTI] - Federal Re- search Institute for Rural Areas, Forestry and Fisheries - Insti- tute of Sea Fisheries Palmaille 9, D-22767 Hamburg, Germany	Tel. +49 40 38905-228 Fax: +49 40 38905-263	christoph.stransky@vti.bund.de

Annex 2: Agenda

Tuesday 16 September 2008

9:00 - 9:30 Start of the meeting

- Housekeeping, network access
- Suggestions for venues of lunch breaks and dinner
- Adoption of the agenda
- Election of the chairman

9:30 – 12:00 Review of logistics and sampling protocols (plenary)

- Area covered, duration of the survey, deviation form survey planning
- Trawling: gear, number and duration of trawls, trawling depths
- Biological sampling: review of the protocol and possible deviations from it.
- Hydroacoustics: transducer, calibration, noise measurements, inter-vessel comparisons
- Hydrography: equipment, precision, sampling protocol
- Daily reporting protocol

12:00 – 13:00 Lunch break

13:00 – 17:00 Review of data analysis protocols (plenary)

- Species composition
- Length frequency distribution, sex-ratio, age distribution
- Parasites
- Acoustics scrutinizing, SA allocation, DSL, TS, echo-counting,
- Abundance estimates above DSL, within DSL and below DSL
- Data formats

Wednesday 17 September 2008

9:00 – 12:00 Data and figures preparation (sub-groups)

- Trawl data table
- Survey plot: cruise track, location of trawls, CTDs, ...
- Population structure: length distribution (by depth/area/sex), sex-ratio (by depth/area)
- Plot of sA by 5nm, trawl catch rates
- Hydrography tables, maps and sections

• Abundance estimates by region and depths

12:00 - 13:00	Lunch break
13:00 - 15:00	Data and figures preparation continues (sub-groups)
15:00 - 17:00	Report on results (plenary)

Thursday 18 September 2008

9:00 – 12:00 Report drafting (sub-groups) 12:00 – 13:00 Lunch break 13:00 – 17:00 Plenary final

- Main results / Executive summary
- Recommendations for future surveys
- Recommendations to ICES/SGRS

17:00 end of the meeting

Annex 3: Recommendations

1) Expansion of survey area

In 2008, not all the geographical distribution of *S. mentella* could be covered by the survey. The group has identified the need to increase the geographical extension of the survey, and in particular to increase the number of vessels by including at least one further participating vessel (EU, Iceland)

2) Research notifications: The requests for permission to fish in different EEZ by different vessels have to be sent to the relevant authorities well in advance (at least 6-9 months before the start of the survey). The omission of survey effort in a particular EEZ due to missing approval of research activities in an EEZ has lead to unnecessary gaps in survey data in the past and must be avoided by any means. As a way forward, the group recommends that the requesting country stays in regular communication with a contact person in the authorising country about the request and reports problems with the research notification to the authorising country and the survey co-ordinator as soon as possible.

3) Need for harmonization of methodology and protocols:

The work of the group was hampered by insufficient harmonisation of instrumentation (hyroacoustics, hydrography), trawl gear and biological sampling. There is an urgent need for workshops dedicated to the harmonization of scrutinisation of hydroacoustic raw data; survey gear; depth zones to be covered by gear; hydrographic equipment; biological sampling.

4) Next surveys

The group recommended that the next survey should take place in August 2009. The following survey will take place in August 2010 and then every two years in alternance with the international survey in the Irminger Sea-

5) Planning of surveys 2009 and 2010 to be conducted by a new Expert Group (cf. PGRS Draft Resolution)

The planning and reporting of the redfish surveys in the Norwegian Sea should be carried out by a specific planning group. For that purpose, it is recommended that a new ICES expert planning group: the Planning Group on Redfish Survey(PGRS), be set up rapidely. A draft resolution is given in Annex 4.

6) More research on in-situ target strength needed

The true value of *S. mentella* target strength (TS) is still debated. Current uncertainty on the true TS value can lead to over-estimates (or under-estimates) of stock abundance by a factor of 2. The group recommended that effort should be put on in situ measurement of redfish target strength.

7) Scrutinisation workshop

Variations in the acoustic performances of the three vessels used in 2008 and in the acoustic scrutinizing methods led to important differences in mean abundance. To resolve the issue, it <u>wais</u> proposed that a workshop on the analysis of hydroacoustics raw data <u>beis</u> hosted rapidly. Proposal: 3 days, late Nov. 2008, in Tromsø, Norway. The workshop was help in Tromsø, Norway in November 2008.

8) Maturity staging workshop

The maturity data from Russia, Norway and the Faroes revealed some inconsistency. The group recommended that a workshop on maturity reading be hosted to resolve methodological differences. This workshop will be proposed to ICES PGCCDBS (Planning Group on Commercial Catch, Discards and Biological Sampling) in 2009 and if approved it will be conducted in 2010

9) Otolith exchange and mini-workshop

The group recommended that harmonisation of otolith age readings should be carefully considered. Exchange of otoliths and the setting up of a mini-workshop on otolith reading are proposed, within the existing bilateral collaboration program between Russia and Norway. Countries other than Norway and Russia will be invited.

Annex 4: Recommendation for a new planning group

The **Planning Group on Redfish Surveys** [PGRS] (Chair: Benjamin Planque, Norway) will be established and takes place at ICES Headquarters, Copenhagen, from 28-30 January 2009, and in Bergen, Norway, from 1-3 September 2009 to:

- a) plan the international trawl/acoustic survey on redfish in the Norwegian Sea and adjacent waters in August 2009 (January meeting);
- b) prepare the report on the outcome of the 2009 survey (September meeting);
- c) consider the establishment of an international database for redfish surveys in the Norwegian Sea. $\,$
- PGRS will report by 15 March 2009 (January meeting) and 15 October 2009 (September meeting) for the attention of the Resource Management Committee, Arctic Fisheries Working Group, Study Group on Redfish Stocks and ACOM.

Supporting Information

Supporting Infor	mation
PRIORITY:	Essential
SCIENTIFIC JUSTIFICATION AND RELATION TO ACTION PLAN:	PGRS will be responsible for the planning and reporting of the international hydroacoustic-trawl survey on pelagic redfish (<i>Sebastes mentella</i>) in the Norwegian Sea and adjacent waters. Redfish in this area have been fished in an olympic fishery by an international fleet since 2005. Since 2007, ICES has advised a protection of juveniles, no directed trawl fishery and low bycatch limits for <i>S. mentella</i> in Sub-areas I and II. NEAFC has recently set a TAC for pelagic <i>S. mentella</i> in this area of 14,500 t. The unknown stock size and its relations to other <i>S. mentella</i> stocks on the shelves have evoked the immediate need for an international survey on redfish in the Norwegian Sea and adjacent waters. The first international survey, carried out in August 2008, was hampered by insufficient harmonisation of instrumentation (hyroacoustics, hydrography), trawl gear and biological sampling. The need for a planning group on redfish surveys in the Norwegian Sea, with close linkage to the Study Group on Redfish Stocks (SGRS) was clearly identified in the post-survey meeting of the 2008 survey. Moreover, the expansion of the survey area and participation of more than three vessels has been recommended. From the early stages of the survey, it is highly advisable to build up an international database for redfish surveys in the Norwegian Sea, including scrutinised hydroacoustic data, biological data and hydrographic data. Relation to Strategic Plan: Provide sound, credible, timely, peer-reviewed, and integrated scientific advice on fishery management and the protection of the marine environment in response to requests from regulatory commissions, Member Countries, and partner organisations.
RESOURCE REQUIREMENTS:	N/A
PARTICIPANTS:	<10 (incl. the cruise leaders of each vessel and the principle experts involved in abundance and biomass calculations). Participation of SGRS members is highly recommended due to the expected synergistic effects in the planning of the survey and analysis of hydroacoustic, biological and hydrographic data.
SECRETARIAT FACILITIES:	N/A
FINANCIAL:	Travel costs will be eligible for participants from Member States of the European Union throgh the EU Data Collection Regulation (Reg. 199/2008).
LINKAGES TO ADVISORY COMMITTEES:	ACOM
LINKAGES TO OTHER COMMITTEES OR GROUPS:	RMC, AFWG, SGRS, PGNAPES
LINKAGES TO OTHER ORGANISATIONS:	NEAFC

Annex 5: Planning document for the *Sebastes mentella* survey in the Norwegian Sea in August 2008

Objective of the survey:

The objective of the survey is defined in the NEAFC document AM 2007/58 which states that the Contracting Parties agreed to conduct a scientific survey for pelagic *Sebastes mentella* in ICES Sub-areas I and II during August-September 2008 to measure the horizontal and vertical stock distribution and provide an abundance estimate. The survey planning (below) has been designed to fulfil these objectives.

Spatial and temporal extent of the survey:

To conform to NEAFC request, the proposed survey design covers the known distribution area of *S. mentella* in the Norwegian Sea (as observed through past scientific surveys and commercial catches). Originally, the survey plan involved 5 vessels (one by contracting parties: Russia, Iceland, Faeroes, EU and Norway) for a duration of 15 days. The survey is planned to start on the 11th August (Monday). Soon after the start of the survey the vessels will be located in the central part of the survey area. A cross-vessel calibration exercise will then be conducted. Map of the planned cruise tracks are given in Appendix 1.

It has been planned that the survey will be funded by research quotas. Possibly up to 2000 tons, i.e. 400 tons per participating parties. However, the attribution of research quotas in international waters and the use of these quotas by contracting parties is still to be clarified. Norway will guarantee that the remaining tons after the scientific survey may be caught in a directed fishery within the Norwegian Economic Zone (incl. Jan Mayen) or the Svalbard Fishery Protection Zone. All participating parties that will depend on such a guarantee should as soon as possible apply the Norwegian authorities for permission to enter the NEZ and/or Svalbard as part of the research survey, incl. permission to fish up to 400 t in these zones together. Vessels participating to the survey will communicate daily. The procedure for communication will be defined when the participating vessels have been identified.

Iceland and EU will not be able to operate a survey vessel in 2008. The survey design presented in Appendix 1 is based on participation from three vessesl: Norway, Russia and the Faeroes.

Acoustic and trawling:

To fulfil the primary objective of the survey (above) the sampling protocol will be based on acoustics transects combined with pelagic trawl hauls. This methodology is adapted to resolve the horizontal and vertical distribution of redfish within the first 500m and possibly deeper. It is currently believed that off-shelf, most of the *S. mentella* population in the Norwegian Sea is located in the 300-500m depth layer. The combined acoustics-trawl survey should therefore form the basis of stock assessment in the area.

However, as the presence of *S. mentella* in deeper layers cannot be excluded, additional deep 'blind' fishing will be conducted during the survey. This will consists of trawl hauls performed at depth greater than 500m and possibly down to 1000m.

Although Faeroes has expressed some difficulties to operate an acoustics survey, it is willing to achieve such protocol. Iceland has offered personnel and expertise for the

Faeroese component of the survey, if needed. For each participant, the cruise track approximates 2000 nautical miles and 37 trawl hauls, cruise tracks are provided in Appendix 1. The first day of the survey should be used for acoustics calibration. Acoustics cross-calibration between vessels is planned at the beginng of the survey between Russian and Norwegian vessels and at the end of the survey, between Russian and Faeroese vessels. Due to the geographical configuration of the cruise track, there will be no direct calibration between Faeroese and Norwegian vessels.

Technical specifications for acoustics

The specifications for acoustics are provided in Appendix 2. There is consensus on using split beam transducer and 38KHz as the primary frequency. Particular attention should be paid to vessel acoustics characteristics when selecting the operating fishing vessels.

Technical specifications for trawls

The fishing activity during the time of the survey will be restricted to direct contributions to the primary objective (i.e. measure the horizontal and vertical stock distribution and provide an abundance estimate), whether participating vessels are allocated specific research quotas or not.

Trawl hauls will be used for species and size composition in conjunction with acoustics so there is no need for an exact match of fishing gears between vessels. However, to limit possible discrepancies in catch characteristics between vessels it was decided to favour the common use onboard all vessels of a Gloria trawl 2048 of not less than 100 mm mesh size, with a 12m cod-end fitted with an inner net of 40mm mesh size, or an equivalent trawl (see Appendix 3). The Norwegian survey will use a multisampler cod-end in order to resolve fish depth distribution with more accuracy. As this equipment is not available to other countries, the trawls should be integrated over sufficient depth. Trawl hauls will be set to last approximately 2 hours (+1h handling).

Calibration & Inter-calibration

The first day of the cruise will be required to perform acoustics calibration. This will be done using a standard sphere calibration procedure. This calibration is essential to ensure the quality of the acoustics data collected. The calibration should be repeated (e.g. at the end of the survey) in case of any doubt. At the beginning of the survey all vessels will be heading for the same geographical area and close to the area of greater density of *S. mentella*. Acoustics cross-calibration between vessels will then be conducted, i.e. with vessels steaming in parallel and acquiring acoustics data simultaneously^{1,2}.

Biological sampling

Biological sampling should be conducted with the objective of: (i) quantifying species composition in the catch, (ii) resolving sex, age, length, maturity, and parasite infesta-

¹ Monstad T, Borkin L, Ermolchev V (1992) Report of the joint Norwegian-Russian acoustic survey on blue whiting, spring 1992. ICES CM 1992/H:6

² Section 8.9.2 in Simmonds J, MacLennan (2005) Fisheries Acoustics, theory and practice. 2nd edition. Blackwell Publ. 437pp.

tion of *S. mentella*, and (iii) collecting biological material for subsequent genetic analysis of *S. mentella*.

Key points for sampling:

- Sub-sampling: In the case of sub-sampling, the ratio of the sub-sample to the total catch should be noted as "conversion factor" in the data recording sheet. Subsampling should be done from the top middle and bottom part of the cod end.
- 2. Catch composition: Taxonomic identification, length and weight will be recorded for all individual fish from the (sub) sample.
- 3. Redfish individual data: The total length (cm below), individual weight, sex and stage of maturity should be measured on at least 100 redfish from each haul. The maturity scale given in Appendix 6 will be used for data exchange. The Russian participants will use the maturity scale given in Appendix 7 that will be converted to the one given in Appendix 6. When using multiple cod ends (Norway) the individual sampling procedure should be carried on each cod end.
- 4. Redfish otolith sampling: A pair of otolith will be collected from randomly selected individual fish sampled for weight, sex and stage, up to a maximum of 30 fish per station. The otolith envelope should carry the station no. and fish ID no. given in the database to allow for allocation to the individual biological data.
- Redfish genetic sampling: Fin clips will be randomly sampled from 100 *S. mentella* from each haul and from all fishes on which ageing is performed.
 Collected samples will be preserved in ethanol for subsequent micro-satellite analysis.
- 6. Redfish parasite and pigments: Presence of parasites (*S. lumpi*) and location of pigment spots will be recorded on individual fish (as in 3). Parisite recording sheet is given in Appendix 8.

Hydrological data collection

Hydrological data will be collected in addition to acoustics and biological data. CTD casts may be the optimal way of acquiring vertical hydrological profiles. However, this would require a specific winch and will take some extra time to deploy during the survey. This may not be achievable. Instead hydrological measurements will be performed using a CTD probe attached to the trawl, and recording depth, temperature and conductivity while trawling. The recommended settings for CTD measurements are: maximum depth range of 1000m, sampling frequency 1Hz or more, temperature accuracy 0.01°C, salinity accuracy 0.02, pressure accuracy 0.02% full scale (i.e. 0.2m for 1000m depth range). Instruments should be calibrated before the start of the survey.

Data format and exchange

Two possible data formats have been proposed, originating from the Irminger Sea surveys (ICES SGRS) or from the Norwegian Sea pelagic surveys (ICES-PGNAPES). The data from SGRS will be used for the survey. The data exchange forms used for the Irminger Sea survey will be used to exchange information between vessels once a day.

Other considerations

Time synchronisation of all instruments used for data collection should be performed at the beginning of the survey and regularly checked. UTC will be the time reference for all data collected.

Geographical positioning of the vessels should be monitored and recorded using a GPS system. Data should be transmitted from the GPS to the echosounder software.

If equipment is available to measure ship angular motion, this should be recorded.

When trawling, available data on altitude, depth and geometry of the trawl should be recorded.

Application to fish in national waters

Following the cruise tracks proposed prior to the London meeting, Russia will have to request permission to enter Norwegian waters for the purpose of the survey. Since Iceland will not take part directly to the survey, Faeroes may wish to extend its survey tracks into Icelandic waters in the case of detectable redfish presence. Faeroes should therefore seek permission to enter Icelandic waters for the purpose of the survey.

Scientists in charge

Benjamin Planque (IMR-Tromsø, benjamin.planque@imr.no) will be responsible for the coordination of the survey and will be in charge of the Norwegian part of the survey.

Andrey Pedchenko (PINRO-Murmansk, andy@pinro.ru) will be in charge of the Russian part of the survey. Dmitriy Aleksandrov (mitja@pinro.ru) will be cruise leader for the Russian part of the survey.

Jákup Reinert (FRS-Tórshavn, jakup@frs.fo) will be in charge of the Faeroese part of the survey.

A group composed of the leading scientists from each party should meet after the survey to exchange on aspects concerning the survey design, data exchange and data processing. This meeting will be held under the auspices of ICES, at ICES headquarters in Copenhagen from 16 to 18 September. The group will report to the ICES Study Group on Redfish Stocks (SGRS).

Vessels

The Russian part of the survey will take part onboard F/V Osveiskoe. Registration K-2165, call sign UDXD. Email: osveisko@plitfor.koenig.ru

The Norwegian part of the survey will take part onboard F/V Atlantic Star. Registration F-111-BD, call sign LMBG. Email: atlantic.star@seamail.no

The Faeroese par of the survey will take part onboard F/V Skálaberg. Registration KG118, call sign XTPI.

Acoustic survey tracks

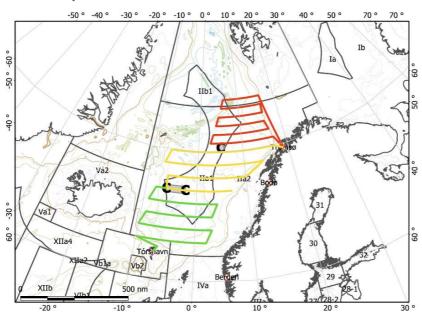


Figure 1: Norwegian Sea redfish acoustic survey sampling plan for 2008. ICES areas are delimited with black lines. Coloured lines indicate bathymetry. Green Track: Faeroes, Red track: Norway, Yellow track: Russia. Grey areas labelled with "C" indicate location of inter-vessel calibration.

Table 1. Total distance, track length and number of trawls estimated for each participating country. The total number of hours required is estimated with vessel steaming at 10 knots, each trawl haul lasting 3h and 8h additional time (for acoustic calibration). Inter-vessel calibration is accounted for.

Country	Total dis- tance	Sampling distance	hauls	Total hours required
Faeroes	1842 nm	1345 nm	32	297
Russia	2001 nm	1595 nm	40	337
Norway	2008 nm	1332 nm	39	335
EU	-	-	-	-
Iceland	-	-	-	-

Acoustics specifications

Survey vessels should be equipped with an echsounder Simrad EK60 or similar. The parameters settings should follow the following specifications:

Transducer: ES38B or equivalent (split beam)

Primary frequency: 38kHz

Pulse length: 1ms Beam angle: 7 degrees Transmit Power: 2kW

Sampling rate $^{\sim}$ 1Hz. This may lead to ocean floor ghost echoes in areas deeper than $^{\sim}$ 750m, in such case, this should be slightly adjusted in a ad hoc fashion to remove ghost echoes from the layer containing redfish. This sampling rate also implies that fish echoes will be only recorded at depth of less than $^{\sim}$ 750m.

Vessel acoustic performance: Each institute should follow the recommendations froms ICES³ when selecting the fisheries vessel which will participate to the survey.

The recommended echosounder is EK60/EK500 or equivalent. If possible it is recommended that a scientific echosounder is brought onboard the vessel and serves as the master echosounder. If several acoustic systems co-exist onboard the vessel, these should be synchronized and the echosounder used for scientific purpose should be configured as master and other equipment as slaves.

Calibration should be performed at the beginning of the survey using a standard calibration sphere, following recommendations from Foote et al⁴. Each parties should ensure that calibration sphere as well as associated equipment and software for the calibration are brought onbard.

The target strength (TS) for *S. mentella* is defined using the following size-dependent relationship: $TS = 20 \log L$ -68. This corresponds to the most recent TS estimate based on measurements performed at close vicinity of individual fish⁵,⁶,⁷. There is still ongoing debate on the most appropriate value of TS to be used for *S. mentella*. This will be further discussed, likely within ICES, and the TS equation will be revised if necessary.

³ ICES (2007) Collection of acoustic data from fishing vessel. ICES Coop. Res. Rep. 287, 84pp.

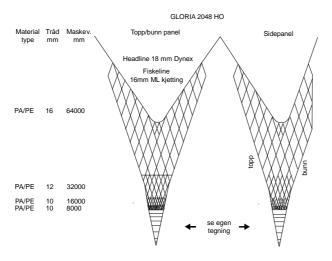
⁴ Foote, K.G., Knudsen, H.P., Vestnes, G., MacLennan, D.N. and Simmonds, E.J. (1987) Calibration of acoustics instruments for fish density estimation: a practical guide. ICES Coop. Res. Rep. 144, 57pp.

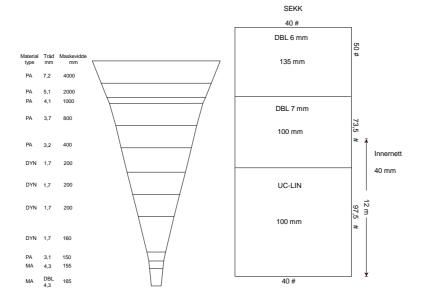
⁵ Gauthier, S. and G. A. Rose (2001) Target Strength of encaged Atlantic redfish (*Sebastes spp.*). ICES Journal of Marine Science 58(3): 562-568.

⁶ Gauthier, S. and G. A. Rose (2002) In situ target strength studies on Atlantic redfish (*Sebastes spp.*). ICES Journal of Marine Science 59(4): 805-815.

⁷ Kang, D. and D. Hwang (2003) Ex situ target strength of rockfish (*Sebastes schlegeli*) and red sea bream (*Pagrus major*) in the Northwest Pacific. ICES Journal of Marine Science 60(3): 538-543.

Trawl specifications





Data format and exchange:

As in SGRS

Annex 5 / Appendix 5

700 800

Horizontal and vertical distribution of redfish *S. mentella* during the Norwegian Sea survey conducted by IMR-Norway in 2007.

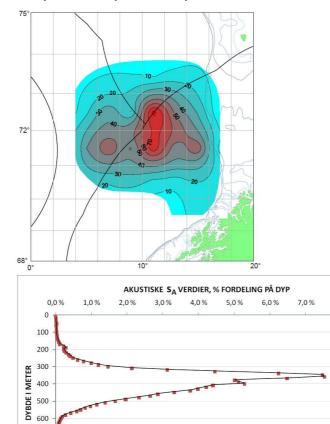


Figure 2. Top: Horizontal distribution of redfish area backscattering coefficient (sA, m²/nmi²) in the area surveyed in 2007. Bottom: vertical distribution of redfish observed by acoustics during the Norwegian Sea survey in August 2007. Vertical axis: depth in meter. Horizontal axis: relative frequency of redfish area backscattering coefficient (sA) per 10m depth layers. Highest redfish densities where observed between 300 and 500m.

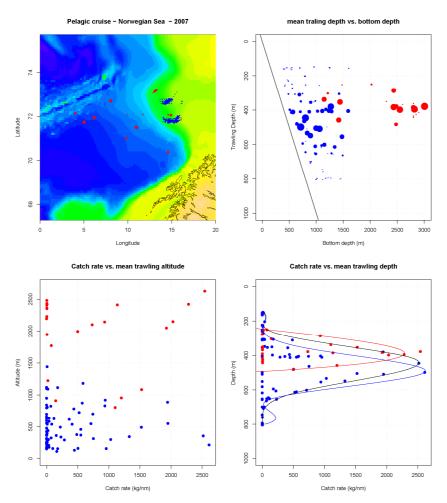


Figure 3. Top-left: spatial distribution of survey trawl hauls in summer 2007. Blue: first period, red: second period. Size of dots is proportional to S. mentella catch rate. Background colour indicates bathymetry. Top-right depth of trawling vs. Bottom depth . Size of dots is proportional to S. mentella catch rate. Bottom-left: vertical distribution of catch rates against trawling altitude. Bottom-right: vertical distribution of catch rates against trawling depth. Lines are the maximum catch rates for the first (blue), second (red) or both (black) periods, fitted using quantile regression splines (τ =0.9).

Maturi ty scale agreed to be used in the international survey in June/July 2007 for redf ish in the Irminger Sea and adjacent waters (reproduced from the report of the ICES study group on redfish stocks, ICES CM 2007/RMC:01

MATURITY STAGES OF FEMALE REDFISH

Stage	Code	Ovaries description
Immature	1 (I)	Ovaries tubular, thin and small. Ovarian wall whitish and delicate. Without conspicuous blood vessels, If visible eggs occur, they are very small, whitish or pale yellowish. Pigmented eye larvae are never observed in the ovary.
Maturing/ Mature	2 (M)	The ovary has increased in size considerably and it is easy to distinguish in the body cavity. The ovary wall and eggs inside the ovary are clearly visible. Eggs are yellow and opaque.
Mature/ Fertilized	3 (F)	Ovaries are considerably bigger and occupy most of the body cavity. Colour is bright yellow. Many eggs are transparent (approx, 50%) because of yolk re-absorption the eye pigment of the larvae becomes visible.
Parturition	4 (P)	Ovary occupy practically the whole body cavity, it is delicate and the wall transparent and thin. The colour shift to a green- yellowish due to larval developing, the eyes are evident and there is little yolk. Larvae are easily released from the ovary when it is manipulated.
Post spawning	5 (S)	Ovary is flaccid, but still big. No visible larvae inside or just a remainder of them. The colour is purple or blackish, sometimes confused with the body cavity wall (peritoneum).
Recovery	6 (R)	Size is reduced to stage 3 or smaller, but no visible eggs, colour yellow to purple.

MATURITY STAGES OF MALE REDFISH

Stage	Code	Testes and genital papilla description
Immature	1 (I)	Testes are translucent, very thin and sometimes even difficult to detect, because it is confused with the mesentery. Width less than 1 mm. The penis is difficult to distinguish and easy to confuse with female genital papilla.
Maturing/ Mature	2 (M)	The testes are more easily distinguishable because of increasing size. They are white. Width more than 1,1-1,5 mm. There is no running sperm when the testes are cut. Penis is visible, and it is easy to identify sex externally.
Mature/ Fertilized	3 (F)	Testes are bright white. The sperm is observed inside the testes, but only when they are cut, i.e. sperm doesn't run out of the testes when they are pressed. Penis is thick, but no sperm is observed on it.
Parturition	4 (P)	Testes are big and with a cream colour. The sperm run out of the fish when belly is pressed. Penis is very conspicuous, with a purple tip and there are remains of sperm on it.
Post spawning	5 (S)	Testes are flaccid. The colour is still cream but with obvious dark (brown) patches. Practically no sperm inside the testes.
Recovery	6 (R)	Size of the testes has been reduced to stage 3, but the sperm is not visible. The colour is whitish.

Maturity scale used by Russia in the international survey in June/July 2007 for redfish in the Irminger Sea and adjacent water (reproduced from the report of the ICES study group on redfish stocks, ICES CM 2007/RMC:01)

	MALES
Juvenile	Gonads are poorly developed, sex is indistinguishable. Specimens at this stage
stage	occur throughout a year.
Stage 1	Sex is distinguishable. Testicles are as thin long colourless bends and occur
	throughout a year.
Stage 2	Testicles are as thick long bends, on a cross section they are of irregular
10 1 Dich	triangular shape of brownish colouring. Remnants of non-extruded sperm are
	available in repetitive-maturing specimens. December-March.
Stage 3	Testicles are large, elastic, coloured brown, in some cases they are of violet
	shade. Along a cross section they are of triangular shape with smoothed angles.
	March-June.
Stage 4	Testicles are large, of light-brown colouring, with a white colour being
	irregular in some areas. At the end of the stage the testicles are white due to the
	sperm formed. Along the cross section the sperm does not run. June-September.
Stage 5	Mating period. Testicles are of milky-white colour, When dissecting the
	external sides flow down and drops of sperm are released from spermatic duct.
	September-November.
Stage 6	Extrusion (after mating). Testicles are of brownish colour with white patches.
	Two zones are visible along a cross section, i.e. brown marginal and white
	middle zones, October-December.
_	FEMALES
Juvenile	
Juvenile stage	
	Gonads are poorly developed, sex is indistinguishable. Specimens at this stage occur all the year round.
stage	Gonads are poorly developed, sex is indistinguishable. Specimens at this stage occur all the year round. Ovaries are poorly developed, of light-yellowish colour, eggs are indistinguishable during a whole year.
stage	Gonads are poorly developed, sex is indistinguishable. Specimens at this stage occur all the year round. Ovaries are poorly developed, of light-yellowish colour, eggs are indistinguishable during a whole year.
stage Stage 1	Gonads are poorly developed, sex is indistinguishable. Specimens at this stage occur all the year round. Ovaries are poorly developed, of light-yellowish colour, eggs are indistinguishable during a whole year. (for repetitive-spawning fish - stage 9-2). Eggs are with 0.2-0.5mm diameter.
stage Stage 1 Stage 2	Gonads are poorly developed, sex is indistinguishable. Specimens at this stage occur all the year round. Ovaries are poorly developed, of light-yellowish colour, eggs are indistinguishable during a whole year. (for repetitive-spawning fish - stage 9–2). Eggs are with 0.2–0.5mm diameter. In immature fish a membrane of ovaries is transparent, in repetitive-spawning specimens it is covered with black pigment. May-August.
stage Stage 1 Stage 2 Stage 3	Gonads are poorly developed, sex is indistinguishable. Specimens at this stage occur all the year round. Ovaries are poorly developed, of light-yellowish colour, eggs are indistinguishable during a whole year. (for repetitive-spawning fish - stage 9–2). Eggs are with 0.2–0.5mm diameter. In immature fish a membrane of ovaries is transparent, in repetitive-spawning specimens it is covered with black pigment. May-August. Ovaries are bright-orange, egg diameter is about 1mm. August-September.
stage Stage 1 Stage 2	Gonads are poorly developed, sex is indistinguishable. Specimens at this stage occur all the year round. Ovaries are poorly developed, of light-yellowish colour, eggs are indistinguishable during a whole year. (for repetitive-spawning fish - stage 9–2). Eggs are with 0.2–0.5mm diameter. In immature fish a membrane of ovaries is transparent, in repetitive-spawning specimens it is covered with black pigment, May-August. Ovaries are bright-orange, egg diameter is about 1mm. August-September. Ovaries occupy above a half of the body cavity, egg diameter is up to 1.5mm.
stage Stage 1 Stage 2 Stage 3	Gonads are poorly developed, sex is indistinguishable. Specimens at this stage occur all the year round. Ovaries are poorly developed, of light-yellowish colour, eggs are indistinguishable during a whole year. (for repetitive-spawning fish - stage 9–2). Eggs are with 0.2–0.5mm diameter. In immature fish a membrane of ovaries is transparent, in repetitive-spawning specimens it is covered with black pigment. May-August. Ovaries are bright-orange, egg diameter is about 1mm. August-September. Ovaries occupy above a half of the body cavity, egg diameter is up to 1.5mm. September-December.
stage Stage 1 Stage 2 Stage 3 Stage 4 Stage 5	Gonads are poorly developed, sex is indistinguishable. Specimens at this stage occur all the year round. Ovaries are poorly developed, of light-yellowish colour, eggs are indistinguishable during a whole year. (for repetitive-spawning fish - stage 9–2). Eggs are with 0.2–0.5mm diameter. In immature fish a membrane of ovaries is transparent, in repetitive-spawning specimens it is covered with black pigment. May-August. Ovaries are bright-orange, egg diameter is about 1mm. August-September. Ovaries occupy above a half of the body cavity, egg diameter is up to 1.5mm. September-December. Ovaries are muddy-greenish, eggs are transparent. December-March.
stage 1 Stage 2 Stage 3 Stage 4	Gonads are poorly developed, sex is indistinguishable. Specimens at this stage occur all the year round. Ovaries are poorly developed, of light-yellowish colour, eggs are indistinguishable during a whole year. (for repetitive-spawning fish - stage 9–2). Eggs are with 0.2–0.5mm diameter. In immature fish a membrane of ovaries is transparent, in repetitive-spawning specimens it is covered with black pigment. May-August. Ovaries are bright-orange, egg diameter is about 1mm. August-September. Ovaries occupy above a half of the body cavity, egg diameter is up to 1.5mm. September-December. Ovaries are muddy-greenish, eggs are transparent. December-March. Ovary membrane is strongly prolonged. The stage lasts from the moment of
stage Stage 1 Stage 2 Stage 3 Stage 4 Stage 5 Stage 6	Gonads are poorly developed, sex is indistinguishable. Specimens at this stage occur all the year round. Ovaries are poorly developed, of light-yellowish colour, eggs are indistinguishable during a whole year. (for repetitive-spawning fish - stage 9–2). Eggs are with 0.2–0.5mm diameter. In immature fish a membrane of ovaries is transparent, in repetitive-spawning specimens it is covered with black pigment. May-August. Ovaries are bright-orange, egg diameter is about 1mm. August-September. Ovaries occupy above a half of the body cavity, egg diameter is up to 1.5mm. September-December. Ovaries are muddy-greenish, eggs are transparent. December-March. Ovary membrane is strongly prolonged. The stage lasts from the moment of cleavage to the beginning of eye pigmentation in embryo. December-March.
stage Stage 1 Stage 2 Stage 3 Stage 4 Stage 5	Gonads are poorly developed, sex is indistinguishable. Specimens at this stage occur all the year round. Ovaries are poorly developed, of light-yellowish colour, eggs are indistinguishable during a whole year. (for repetitive-spawning fish - stage 9–2). Eggs are with 0.2–0.5mm diameter. In immature fish a membrane of ovaries is transparent, in repetitive-spawning specimens it is covered with black pigment. May-August. Ovaries are bright-orange, egg diameter is about Imm. August-September. Ovaries occupy above a half of the body cavity, egg diameter is up to 1.5mm. September-December. Ovaries are muddy-greenish, eggs are transparent. December-March. Ovary membrane is strongly prolonged. The stage lasts from the moment of cleavage to the beginning of eye pigmentation in embryo. December-March. Eye pigmentation begins in embryos owing to which ovaries gradually acquire
stage Stage 1 Stage 2 Stage 3 Stage 4 Stage 5 Stage 6 Stage 7	Gonads are poorly developed, sex is indistinguishable. Specimens at this stage occur all the year round. Ovaries are poorly developed, of light-yellowish colour, eggs are indistinguishable during a whole year. (for repetitive-spawning fish - stage 9–2). Eggs are with 0.2–0.5mm diameter. In immature fish a membrane of ovaries is transparent, in repetitive-spawning specimens it is covered with black pigment. May-August. Ovaries are bright-orange, egg diameter is about 1mm. August-September. Ovaries occupy above a half of the body cavity, egg diameter is up to 1.5mm. September-December. Ovaries are muddy-greenish, eggs are transparent. December-March. Ovary membrane is strongly prolonged. The stage lasts from the moment of cleavage to the beginning of eye pigmentation in embryo. December-March. Eye pigmentation begins in embryos owing to which ovaries gradually acquire black colouring. February-March.
stage Stage 1 Stage 2 Stage 3 Stage 4 Stage 5 Stage 6	Gonads are poorly developed, sex is indistinguishable. Specimens at this stage occur all the year round. Ovaries are poorly developed, of light-yellowish colour, eggs are indistinguishable during a whole year. (for repetitive-spawning fish - stage 9–2). Eggs are with 0.2–0.5mm diameter. In immature fish a membrane of ovaries is transparent, in repetitive-spawning specimens it is covered with black pigment. May-August. Ovaries are bright-orange, egg diameter is about Imm. August-September. Ovaries occupy above a half of the body cavity, egg diameter is up to 1.5mm. September-December. Ovaries are muddy-greenish, eggs are transparent. December-March. Ovary membrane is strongly prolonged. The stage lasts from the moment of cleavage to the beginning of eye pigmentation in embryo. December-March. Eye pigmentation begins in embryos owing to which ovaries gradually acquire black colouring. February-March. Eyes acquire bright metallic shade. Embryos are well developed and mobile.
stage Stage 1 Stage 2 Stage 3 Stage 4 Stage 5 Stage 6 Stage 7	Gonads are poorly developed, sex is indistinguishable. Specimens at this stage occur all the year round. Ovaries are poorly developed, of light-yellowish colour, eggs are indistinguishable during a whole year. (for repetitive-spawning fish - stage 9–2). Eggs are with 0.2–0.5mm diameter. In immature fish a membrane of ovaries is transparent, in repetitive-spawning specimens it is covered with black pigment. May-August. Ovaries are bright-orange, egg diameter is about 1mm. August-September. Ovaries occupy above a half of the body cavity, egg diameter is up to 1.5mm. September-December. Ovaries are muddy-greenish, eggs are transparent. December-March. Ovary membrane is strongly prolonged. The stage lasts from the moment of cleavage to the beginning of eye pigmentation in embryo. December-March. Eye pigmentation begins in embryos owing to which ovaries gradually acquire black colouring. February-March. Eyes acquire bright metallic shade. Embryos are well developed and mobile. The stage lasts until larvae extrusion.
stage Stage 1 Stage 2 Stage 3 Stage 4 Stage 5 Stage 6 Stage 7	Gonads are poorly developed, sex is indistinguishable. Specimens at this stage occur all the year round. Ovaries are poorly developed, of light-yellowish colour, eggs are indistinguishable during a whole year. (for repetitive-spawning fish - stage 9–2). Eggs are with 0.2–0.5mm diameter. In immature fish a membrane of ovaries is transparent, in repetitive-spawning specimens it is covered with black pigment. May-August. Ovaries are bright-orange, egg diameter is about Imm. August-September. Ovaries occupy above a half of the body cavity, egg diameter is up to 1.5mm. September-December. Ovaries are muddy-greenish, eggs are transparent. December-March. Ovary membrane is strongly prolonged. The stage lasts from the moment of cleavage to the beginning of eye pigmentation in embryo. December-March. Eye pigmentation begins in embryos owing to which ovaries gradually acquire black colouring. February-March. Eyes acquire bright metallic shade. Embryos are well developed and mobile.

Sheet for the recording of fish individual data onboard, including parasites and pigmentation.

